

National Academy of Clinical Biochemistry Guidelines: The Use of Microarrays in Cancer Diagnostics

Eleftherios P. Diamandis^{1*}, Manfred Schmitt² and Da-elene van der Merwe¹

¹Department of Pathology and Laboratory Medicine, Mount Sinai Hospital, Toronto, Ontario, M5G1X5, Canada and Department of Laboratory Medicine and Pathobiology, University of Toronto, Toronto, Ontario, M5G 1L5, Canada; ²Clinical Research Unit, Department of Obstetrics and Gynecology, Technical University of Munich, 22 Ismaninger Street, D-81675, Munich, Germany

***Sub-Committee Chair**, to whom all comments should be addressed *via* e-mail to ediamandis@mtsinai.on.ca with a copy to C.Sturgeon@ed.ac.uk

Key words: MALDI-TOF; mass spectrometry; cancer diagnostics

Abbreviations: DNA, deoxyribonucleic acid; FDA, Food and Drug Administration; PCR, polymerase chain reaction; MGED, Microarray Data Standards, Annotations Ontologies and Databases; SNP, Single Nucleotide Polymorphism, TMA, Tissue MicroArray

INTRODUCTION

Microarrays were first commercially introduced in 1996 by Affymetrix (Genechip^R technology) (1). Depending on the biomolecule immobilized on the surface, these devices are known as DNA-chips, protein-chips or cell-chips. Cancer was one of the first areas in which microarrays have been employed for diagnostic purposes. Disease classification, prognosis, monitoring and prediction of therapeutic response are some of the areas where microarrays have the potential to become routine diagnostic tools. This technology enables substitution of linear studies of individual events to parallel and simultaneous analysis of complex systems and pathways. Regardless of the application, the resulting information comprises thousands of individual measurements and provides an intricate and complex snapshot of biological properties of the cell, tissue, organ or fluid.

PRINCIPLES OF MICROARRAYS

A microarray is a compact device that contains a large number of well-defined immobilized capture molecules (e.g. synthetic oligos, PCR products, proteins, antibodies) assembled in an addressable format. The best-known microarrays, DNA-biochips, are miniature arrays of gene fragments attached to a glass or plastic surface. These chips are used to examine gene activity (expression profiling) and identify gene mutations or single nucleotide polymorphisms (SNPs), by hybridization between the sequences on the microarray and a labeled probe (the sample of interest). There are two major methods for microarray fabrication: a) photolithography, as is used in the Affymetrix system (400,000 spots in a 1.25 x 1.25 cm area), and b) mechanical deposition or printing on glass slides, membranes etc. The major advantages of microarrays include: small volume deposition (nanoliters, nL), minimal wasted reagents, access to many genes/proteins simultaneously, massive parallel information, automation and potentially quantification. For more detailed information on microarrays, see specialized books (2, 3) and an entire issue of *Nature Genetics* (4).

TISSUE MICROARRAYS

High-throughput analysis of tissues is facilitated by new technologies such as multi-tissue northern blots, protein arrays or real-time PCR (5-8). However, the problem with these methods is that tissues are disintegrated before analysis, preventing identification of the cell types expressing the gene of interest (9). These and other shortcomings can be overcome by tissue microarray (TMA) techniques (10). TMAs consist of up to a 1000 tiny cylindrical tissue samples (0.6 mm in diameter) assembled on a regular-sized routine histology paraffin block. Sections are cut from TMA blocks using standard microtomes. TMA sections allow simultaneous analysis of up to a 1000 tissue samples in a single experiment. The technique is therefore cost-effective. Despite the small size of arrayed samples, TMA studies generally provide reasonably representative information. TMAs are applied over a broad range of cancer research: prevalence TMAs (11-13), progression TMAs (10, 14-16), prognostic TMAs and TMAs composed of experimental tissues such as cell lines (17,18) or xenografts (19).

APPLICATIONS OF MICROARRAYS

Microarrays have been successfully applied in a variety of settings including

- Gene expression profiling (the most popular application)
- Detection of Single Nucleotide Polymorphisms (SNPs) (Pharmacogenetics)
- Sequencing by hybridization (genotyping/mutation detection)
- Protein expression profiling
- Protein-protein interaction studies
- Whole genome biology experiments

Cancer is a heterogeneous disease in many respects, including its cellularity, different genetic alterations and diverse clinical behaviors. Many analytical methods have been used to study human tumors and to classify patients into groups with similar clinical behavior. Most methods require specialized pathologist interpretation; yet none of the classifications are homogeneous enough. It has been hypothesized that the genetic heterogeneity and clinical behavior of cancer

could be better assessed by studying genome-wide gene expression profiles by microarrays (20). Although the potential of microarrays is yet to be fully realized, these tools have shown great promise in deciphering complex diseases, including cancer (20). A partial list of applications of microarrays in cancer is presented in Table 1 (21-43). As is the case with many new technologies, microarrays have many shortcomings, briefly discussed in the following section.

LIMITATIONS OF MICROARRAYS

Microarray technologies are still evolving and this presents difficulties for standardization and consensus development. There are no 'gold standards' such as reference reagents or bioinformatics algorithms. These standards are essential for comparison of data between laboratories and on different platforms (44). Recent reports suggest that microarray data are noisy and not reproducible (45,46). Furthermore, bias poses a significant threat to the validity of data generated by such technologies (47).

MICROARRAYS: NACB RECOMMENDATIONS

There is little doubt that microarrays will eventually become routine diagnostic tools, and the first commercial devices are already on the market (Table 2). However, this is still a relatively new technology and several parameters need to be further optimized and validated prior to their implementation into routine clinical practice, including: selection of optimal capture molecules, standardized hybridization protocols and standardized data collection and interpretation. For DNA and protein microarrays to be reliable tools, they must possess probe sequences that hybridize with high sensitivity and specificity, thereby allowing specific detection of their intended targets. Results must become more reproducible, robust and interchangeable between laboratories, and quality control and quality assurance systems must be established (44). Determining the appropriate level of analytical and clinical validation needed for each application raises new challenges for scientists in industry, academia and regulatory agencies (48).

Two important issues need to be considered when evaluating microarray expression data

1. Whether the results are valid or accurate for the particular biological system under study, and

2. Whether the data fundamentally describe the phenomenon being investigated (49).

Introduction of artifacts is possible at any time during an array experiment, therefore each component of the procedure must be carefully considered. The validation process can be divided into three areas: experimental quality control, independent confirmation of data and universality of results (49). Furthermore, before implementation of microarrays into routine practice, it will be preferable to automate the process to minimize variability and increase robustness. Array production, like any other diagnostic device, must meet minimum criteria set by the Food and Drug Administration (FDA) (50). The International Meeting on Microarray Data Standards, Annotations Ontologies and Databases (MGED) focuses on standardization of biochips and proposes appropriate guidelines (51, 52). Despite widespread applications of microarrays in research, the level of evidence of these studies for clinical application, as described by Hayes et al (53), is Level V (evidence from small pilot studies that estimate distribution of marker levels in sample population). Based on the information above, the NACB Panel has formulated the recommendations outlined in Table 3.

REFERENCES

1. Lockhart DJ, Dong H, Byrne MC, Follettie MT, Gallo MV, Chee MS, Mittmann M, Wang C, Kobayashi M, Horton H, Brown EL. Expression monitoring by hybridization to high-density oligonucleotide arrays. *Nat Biotechnol* 1996; 14: 1675-80.
2. Bowtell D, Sambrook J, eds. *DNA Microarrays: A Molecular Cloning Manual*, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, New York, 2003.
3. Schena M, ed. *Microarray Analysis*, New Jersey; John Wiley & Sons, Inc., Hoboken, 2003.
4. *Nat Genet Suppl* 2002; 32:461-552.
5. Belin D. The use of RNA probes for the analysis of gene expression. Northern blot hybridization and ribonuclease protection assay. *Methods Mol Biol* 1998;86: 87-102.
6. Kallioniemi OP. Biochip technologies in cancer research. *Ann Med* 2001; 33: 142-47.
7. Bichsel VE, Liotta LA, Petricoin EF III. Cancer proteomics: from biomarker discovery to signal pathway profiling. *Cancer J* 2001; 7: 69-78.
8. Walker NJ. Real-time and quantitative PCR: applications to mechanism-based toxicology. *J Biochem Mol Toxicol* 2001; 15: 121-27.
9. Simon R, Mirlacher M, Sauter G. Tissue microarrays in cancer diagnosis. *Expert Rev Mol Diagn* 2003; 3: 421-30.
10. Kononen J, Bubendorf L, Kallioniemi A. Tissue microarrays for high-throughput molecular profiling of hundreds of specimens. *Nat Med* 1998; 4: 844-47.
11. Garcia JF, Camacho FI, Morente M. Hodgkin's and Reed-Sternberg cells harbor alterations in the major tumor suppressor pathways and cell-cycle checkpoints: analyses using tissue microarrays. *Blood* 2002; 12: 12.
12. Hedvat CV, Hagde A, Chaganti RS. Application of tissue microarray technology to the study of non-Hodgkin's and Hodgkin's lymphoma. *Hum Pathol* 2002; 33: 968-974.

13. Tzankov A, Zimpfer A, Lugli A. High-throughput tissue microarray analysis of G1-cyclin alterations in classical Hodgkin's lymphoma indicates overexpression of cyclin E1. *J Pathol* 2003; 199: 201-7.
14. Richter J, Wagner U, Kononen J, Fijan A, Bruderer J, Schmid U High throughput tissue microarray analysis of cyclin E gene amplification and overexpression in urinary bladder cancer. *Am J Pathol* 2000; 157: 787-794.
15. Bubendorf L, Kolmer M, Kononen J. Molecular mechanisms of hormone therapy failure in human prostate cancer analyzed by a combination of cDNA and tissue microarrays. *J Natl Cancer Inst* 1999; 91: 1758-64.
16. Bubendorf L, Kononen J, Koivisto P. Survey of gene amplifications during prostate cancer progression by high-throughput fluorescence in situ hybridization on tissue microarrays. *Cancer Res* 1999; 59: 803-6.
17. Simon R, Struckmann K, Schraml P. Amplification pattern of 12q13-q15 genes (MDM2, CDK4, GL1) in urinary bladder cancer. *Oncogene* 2002; 21: 2476-83.
18. Hoos A, Cordon-Cardo C. Tissue microarray profiling of cancer specimens and cell lines: opportunities and limitations. *Lab Invest* 2001; 81: 1331-38.
19. Bubendorf L, Kolmer M, Kononene J. Hormone therapy failure in human prostate cancer: analysis by complementary DNA and tissue microarrays. *J Natl Cancer Inst* 1999; 91: 1758-64.
20. Chung CH, Bernard PS, Perou CM. Molecular portraits and the family tree of cancer. *Nat Gen* 2002; 32: 533-540.
21. Lakhani SR, Ashworth A. Microarray and histopathological analysis of tumours: the future and the past? *Nat Rev Cancer* 2001; 1: 151-7.
22. Günther K, Merkelbach-Bruse S, Kwaku Amo-Takyi B, Handt S, Schröder W. Differences in genetic alterations between primary lobular and ductal breast cancers detected by comparative genomic hybridization. *J Pathol* 2001; 195: 40-47.

23. Alizadeh AA, Ross DT, Perou CM, van de Rijn M. Towards a novel classification of human malignancies based on gene expression patterns. *J Pathol* 2001; 195: 41-52.
24. Rubin MA. Use of laser capture microdissection, cDNA microarrays, and tissue microarrays in advancing our understanding of prostate cancer. *J Pathol* 2001;195; 80-86
25. Perou CM, Sorlie T, Eisen MB, van de Rijn M, Jeffrey SS, Rees CA, et al. Molecular portraits of human breast tumours. *Nature* 2000; 406: 747-52.
26. van de Vijver MJ, He YD, van't Veer LJ, Dai H, Hart AA, Voskuil DW, Schreiber GJ, Peterse JL, et al. A gene-expression signature as a predictor of survival in breast cancer. *New Engl J Med* 2002; 347: 1999-2009.
27. Sorlie T, Perou CM, Tibshirani R, Aas T, Geisler S, Johnsen H, et al. Gene expression patterns of breast carcinomas distinguish tumor subclasses with clinical implications. *Proc Natl Acad Sci USA* 2001; 98: 10869-74.
28. Khan J, Wei JS, Ringner M, Saal LH, Ladanyi M, Westermann F, et al. Classification and diagnostic prediction of cancers using gene expression profiling and artificial neural networks *Nat Med* 2001; 7: 673-97.
29. Allander SV, Nupponen NN, Ringner M, Hostetter G, Maher GW, Goldberger N, et al. Gastrointestinal stromal tumors with KIT mutations exhibit a remarkably homogeneous gene expression profile. *Cancer Res* 2001;61:8624-8628
30. Su AI, Welsh JB, Sapinoso LM, Kern SG, Dimitrov P, Lapp H, et al. Molecular classification of human carcinomas by use of gene expression signatures. *Cancer Res* 2001; 61: 7388-93
31. Singh D, Febbo PG, Ross K, Jackson DG, Manola J, Ladd C, et al. Gene expression correlates of clinical prostate cancer behaviour. *Cancer Cell* 2002; 1: 203-9.
32. LaTulippe E, Satagopan J, Smith A, Scher H, Scardino P, Reuter V, et al. Comprehensive gene expression analysis of prostate cancer reveals distinct transcriptional programs associated with metastatic disease. *Cancer Res* 2002; 62: 4499-4506.

33. Takahashi M, Rhodes DR, Furge KA, Kanayama H, Kagawa S, Haab BB, et al. Gene expression profiling of clear cell renal cell carcinoma: gene identification and prognostic classification. *Proc Natl Acad Sci USA* 2001; 98:9754-59.
34. Garber ME, Troyanskaya OG, Schluens K, Petersen S, Thaesler Z, Pacyna-Gengelbach M, et al.. Diversity of gene expression in adenocarcinoma of the lung. *Proc Natl Acad Sci USA* 2001;98: 13784-89.
35. Bhattacharjee A, Richards WG, Staunton J, Li C, Monti S, Vasa P, Ladd C, et al. Classification of human lung carcinomas by mRNA expression profiling reveals distinct adenocarcinoma subclasses. *Proc Natl Acad Sci USA* 2001; 98: 13790-95
36. Dhanasekaran SM, Barrette TR, Ghosh D, Shah R, Varambally S, Kurachi K, et al.. Delineation of prognostic biomarkers in prostate cancer. *Nature* 2001; 412: 822-826
37. Rosenwald A. The use of molecular profiling to predict survival after chemotherapy for diffuse large B cell lymphoma. *New Engl J Med* 2002; 346: 1937-47.
38. Alizadeh AA, Eisen MB, Davis RE, Ma C, Lossos IS, Rosenwald A, et al. Distinct types of diffuse large B-cell lymphoma identified by gene expression profiling. *Nature* 2000; 403: 503-11.
39. Hedenfalk I, Duggan D, Chen Y, Radmacher M, Bittner M, Simon R, et al. Gene expression profiles in hereditary breast cancer. *New Engl J Med* 2001; 344; 539-48.
40. Jazaeri AA, Yee CJ, Sotiriou C, Brantley KR, Boyd J, Liu ET. Gene expression profiles of BRCA-1linked, BRCA-2-linked, and sporadic ovarian cancers. *J Natl Cancer Inst* 2002; 94: 990-1000.
41. Zajchowski DA, Bartholdi MF, Gong Y, Webster L, Liu HL, Munishkin A, et al. Identification of gene expression profiles that predict the aggressive behavior of breast cancer cells. *Cancer Res* 2001;61:5168-78
42. Beer DG, Kardia SL, Huang CC, Giordano TJ, Levin AM, Misek DE, et al. Gene expression profiles predict survival of patients with lung adenocarcinoma. *Nat Med* 2002; 8: 816-24.

43. Welsh JB, Zarrinkar PP, Sapinoso LM, Kern SG, Behling CA, Monk BJ, et al. Analysis of gene expression profiles in normal and neoplastic ovarian tissue samples identifies candidate molecular markers of epithelial ovarian cancer. *Proc Natl Acad Sci USA* 2001; 98: 1176 - 1181
44. Petricoin EF, Hackett JL, Lawrence LJ, Puri RK, Gutman SI, Chumkov K, et al. Medical applications of microarray technologies: a regulatory science perspective. *Nat Gen Suppl* 2002; 32: 474-79.
45. Marshall E. Getting the noise out of gene arrays. *Science* 2004; 306: 630-1.
46. Piccart M, Loi S, van't Veer L, Saghatchian-d' Assignies M, Glass A, Ellis P. Multi-center external validation study of the Amsterdam 70-gene prognostic signature in node negative untreated breast cancer: are the results still outperforming the clinical-pathological criteria? Abstract presented at San Antonio Breast Cancer Symposium <http://www.abstracts2view.com/sabcs/search.php?queryxxxxxx=Piccart&where=authors&intMaxHits=10&search=do>.
47. Ransohoff DF. Bias a threat to validity of cancer-molecular marker research. *Nat Rev Cancer* 2005; 5: 142-9
48. Eisen M, Spellman PT, Brown PO, Botstein D. Cluster analysis and display of genome-wide expression patterns. *Proc Natl Acad Sci USA* 1998; 98: 14863-68.
49. Chuaqui RF, Bonner RF, Best CJM, Gillepsie JW, Flaig MJ, Hewitt SM, et al. Post-analysis follow-up and validation of microarray experiments. *Nat Gen Suppl* 2002; 32: 509-14.
50. Toder R. DNA arrays as diagnostic tools in human healthcare. *Expert Rev Mol Diagn* 2002; 2: 422-28.
51. Larson K. DNA Microarrays; The essential technology. 2001. www.pharmabriefing.com/
52. European Bioinformatics Institute/MGED/Annotations-wg/annotations-wg.html.

53. Hayes DF, Bast RC, Desch CE, Fritsche H Jr, Kemeny NE, Jessup JM, et al. Tumor marker utility system: a framework to evaluate clinical utility of tumor markers. *J Natl Cancer Inst* 1996; 88: 1419-20.

Table 1. Microarray applications in cancer diagnostics

Microarray Technology	Application	Cancer	Reference
Comparative genomic hybridization	Classification	Breast	21, 22
cDNA tissue expression profiling	Classification	Breast	23
	Therapeutic response	Lymphoma	23,24
	Molecular profiling	Prostate	25
Gene expression profiling	Prognosis	Breast	26,27
	Classification	Breast	26,28
	Diagnosis	Ewing sarcoma	29
	Diagnosis	Rhabdomyosarcoma	29
	Diagnosis	Burkitt lymphoma	29
	Diagnosis	Neuroblastoma	29
	Diagnosis	GI tumor	30
	Diagnosis	Prostate	31
	Prognosis	Prostate	32,33
	Diagnosis	Bladder	31
	Treatment tailoring	Breast	31
	Classification	Colorectal	31
	Classification	Gastroesophageal	31
	Classification	Kidney	31
	Prognosis	Kidney	34
	Classification	Ovarian	31
	Classification	Pancreas	31
	Classification	Lung	31,35,36
	Molecular profiling	Prostate	37
	Development stages	B-cell lymphomas	38,39
Mutations	BRCA 1 (breast, ovarian)	27,40,41	
Prognostic signature	Prognosis	Breast	41
		Lung	42
Genome mining	Biomarker discovery	Ovarian	43

Table 2: Some commercially available cancer diagnostic devices based on microarray technology.

Name	Intended Use	Manufacturer
1. Amplichip CYP450	Identifies variations in genes CYP2D6 and CYP2C19 for pharmacogenomics	Roche (www.roche.com)
2. GeneChip Mapping 100K	Whole genome SNP analysis (100,000 SNPs) for establishing disease predisposition	Affymetrix (www.affymetrix.com)
3.MammaPrint CupPrint	70-gene signature for breast cancer prognosis Identifying the primary tumor	Agendia (www.agendia.com)
4.p53 GeneChip	Sequencing of p53 gene for identifying mutations	Affymetrix
5. Tumor PSA Array Tumor Monitoring Array Colorectal Cancer DNA Array cDNA Expression Array	tPSA, fPSA, CEA CEA, AFP, hCG, CA19-9, CA125, CA15-3 TP-53, APC, K-ras, BRAF Ovarian, Breast cancer	Randox (www.randox.com)

Table 3. NACB Recommendations for use of Microarrays in Cancer Diagnostics

-
1. Gene expression microarrays are new and promising devices used for cancer diagnosis, prognosis, prediction of therapeutic response, monitoring and selection of therapy. The level of evidence from published studies, according to Hayes et al. (53) is Level V [lowest category]. Consequently, microarrays should continue to be used as research devices, but not as tools for making clinical decisions.
 2. Standardization and clinical validation of expression microarrays is warranted.
 3. Quality control and quality assurance programs for expression microarrays need to be further developed.
 4. Microarray automation is encouraged for improving reproducibility, throughput and robustness.
 5. Tissue microarrays are devices suitable for high-throughput analysis of large numbers of samples and are recommended for use in clinical trials and retrospective studies for evaluating and validating new tumor markers by immunohistochemistry.
 6. Use of microarrays for single nucleotide polymorphism (SNP) analysis is recommended for establishing haplotypes and for correlating these haplotypes to disease predisposition.
 7. Microarrays are recommended to be used for high-throughput genotyping and mutation/sequence variation detection for cancer diagnostics and pharmacogenomics. More validation is necessary to ensure equivalent results between standard technologies (such as DNA sequencing) and microarray analysis.
 8. Protein microarrays and other similar technologies are recommended to be used as research tools for multiparametric analysis of large numbers of proteins. The level of evidence is not as yet high enough for clinical applications.
-